

# Changes in the Mid gut enzyme activity in the fifth instar larvae of silkworm, *Bombyx mori* (L) fed with mulberry leaves treated with aqueous solution of seed powder of *Syzigium cumini* (L) [(Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]).

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## ABSTRACT

Four different concentrations (10.0 ppm; 20.0 ppm; 40.0 ppm & 50.0 ppm) of the aqueous solution of seed powder of *Syzigium cumini* (L) concentrations was used to treat the leaves of mulberry & fed to the fifth instar larvae of bivoltine, crossbreed silkworm, *Bombyx mori* (L) [(Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27)] for first four days of fifth instar larvae. The larvae fed with untreated & water treated leaves were also maintained. The midgut enzyme (protease & amylase) bioassays were carried out on fifth day. The velocity of biochemical reaction catalyzed by mid gut protease and midgut amylase in larvae fed with untreated mulberry leaves was found measured 02.593 units and 5.547 units respectively. The midgut protease activity in larvae fed with mulberry leaves treated with various concentrations (10.0 ppm; 20.0 ppm; 40.0 ppm & 50.0 ppm) of the aqueous solution of seed powder of *Syzigium cumini* (L) was found measured 3.217; 4.339; 4.476 and 5.793 units respectively. There was 24 to 123 percent increase in the mid gut protease activity through *Syzigium* treatment. The midgut amyase activity in experimental group larvae in attempt was found measured 6.864; 10.148; 10.319 and 10.483 units respectively. There was 23 to 88 percent increase in the mid gut protease activity through *Syzigium* treatment. The contents of seed powder of *Syzigium cumini* (L) serve to improve the digestibility & exert the influence of efficient metabolism in the fifth instar larvae of silkworm, *Bombyx mori* (L). The *Syzigium* seed powder treatment may gear overall biochemical constituency of silkworm larvae, through the significant improvement in the velocity of mid gut enzyme catalyzed biochemical reactions.

**Keywords :** *Bombyx mori* (L); Midgut Protease; Midgut Amylase; *Syzigium cumini* (L).

## I. INTRODUCTION

The tree was introduced to Florida in 1911 by the USDA, and is also now commonly grown in Suriname, Guyana and Trinidad and Tobago. In Brazil, where it was introduced from India during Portuguese colonization, it has dispersed spontaneously in the wild in some places, as its fruits are eagerly sought by various native birds such as thrushes, tanagers and the great kiskadee. This species is considered an invasive

in Hawaii. There is close interlinking between the life of insect herbivores and metabolites in plants. The metamorphosis in insects is said to be in the orchestrate progression. There is close interlinking between the life of insect herbivores and pmetabolites in plants. The metamorphosis in insects is said to be in the orchestrate progression. The insect metamorphosis is closely interlinked with plant metabolites. According to Bowers, et al (1966) the chemical constituents of plants (Roots; Stems; Leaves and Fruits)

could have been the factors of growth & metamorphosis for insects. The plant eating insects are able to avoid poor quality food. That is to say, the insects are able to select food from variety available for them. The larvae of silkworm, *Bombyx mori* (L) are monophagous. They are feeding exclusively on the leaves of mulberry *Morus alba* (L). For the purpose of getting qualitative silk cocoons, it is essential to fortify either the quality of food (mulberry leaves) appetite of larval instars of silkworm, *Bombyx mori* (L). According to Murugan and George (1992), the factors responsible for influencing the growth, development & subsequent physiology of body of silkworm larvae include: quality of nutrition, that is to say the biochemical status of nutrients in the food (Leaves of mulberry, *Morus alba* L ); quantity of hormones (hormonal level) in the body & the conditions of climate (environmental conditions). Each and every element in body of larva is primarily derived from it's source of food material. The leaves of mulberry, *Morus alba* (L) are exclusive source of nutrients for the life of larval instars of silkworm, *Bombyx mori* (L). The leaves of mulberry, *Morus alba* (L) are containing the nutrients and many stimulants for the life of larval instars of silkworm, *Bombyx mori* (L) ( Ito, 1960,1961; Nayar & Fraenkel, 1962; Ito, et al, 1964; Ito & Hyashiya, 1965). The quality of the nutrition ( leaves of mulberry, *Morus alba* L.) serves a lot to accelerate the growth, metamorphosis in larval instars of silkworm, *Bombyx mori* (L). The entire credit of life of silkworm, *Bombyx mori* (L) goes to the nutrients in the leaves of mulberry, *Morus alba* (L). Therefore, the leaves of mulberry, *Morus alba* (L) forms the physiological foundation for sericulture. The leaves of mulberry are the mulberry, *Morus alba* (L). The leaves of mulberry, *Morus alba* (L) biochemically constituted with proteins, lipids, carbohydrates (Murali, 1992) & minerals (Subramanyam Reddy, 1992). The biochemical profile of the leaves of mulberry, *Morus alba* (L.) exert influence on the corresponding diversity of larval mid-gut enzymes capable of hydrolyzing the biocompounds in the body of larval instars of

silkworm, *Bombyx mori* (L). The proteins; lipids; carbohydrates (glycogen) are stored in the body tissues of larval instars of silkworm, *Bombyx mori* (L) especially, the fat bodies.

There is variation in the food consumption in phytophagous insects. This may be for varied biochemical processes, ultimately for successful adaptations ( Slansky, 1982). It has been suggested that, there is a functional difference between the activity of digestion by the digestive fluid in mid gut & tissue of mid gut. It has been reported by Horie, et al (1963) that, molecular proteins are hydrolyzed into peptides by digestive fluid content & into aminoacids with peptidases in the mid gut tissue. Likewise, the polysaccharides, are digested in the insect gut lumen by digestive fluid & disaccharides and/or trisaccharides get hydrolysed into their constituent monosaccharide sugars mainly in the gut tissue (Horie, 1967). Yamafuji and Yonezawa (1935) reported the analogy of insect lipase, the lipid digesting enzyme of the insect mid gut with pancreatic lipase of vertebrates. The attempts towards production of the qualitative silk through the improvement in the efficiency of consumption & utilization of food by larval instars of silkworm, *Bombyx mori* (L) include: improvement in the quality of mulberry leaves & supplementation of nutrient biocompounds like soya protein; potassium iodide, copper sulphate, other mineral salts, herbal products ( or drugs) like digoxin (Vitthalrao & Kulkarni, 2011) kho-go ( Desai, et al, 2011) and stevia inulin (Shubhangi Pawar, et al, 2017). Quality of mulberry leaves get reflected into the quality of the cocoons spun by fifth instar larvae of silkworm, *Bombyx mori* (L). There are reports on Use of soya protein; potassium iodide, copper sulphate, mineral salts, herbal products for improvement of the quality of leaves of mulberry, *Morus alba*. Herbal products are well known for the acceleration of metabolism in the body of larval instars of silkworm, *Bombyx mori* (L).

Sericultural practices are basically related to the nutrition and physiology of digestion in silkworm. Moreover, nutrition and physiology of digestion in silkworm are the most fundamental and important challenges in the sericulture. Significant sericulture may occur if and only if a species of silkworm can be grown quickly and economically. Distinguishing feature of larval instars of silkworm is digestion of albumin, fat and carbohydrates except cellulose (Kellner, et al, 1887). The nutrient composition of the meal get reflect on ability of secretion of digestive enzymes in larval instars of silkworm. The leaves of mulberry, *Morus alba* (L) should be supplemented with various nutrients. This may help for silkworm feeding to promote silk quality and quantity (Mahmood et al, 2002). Studies carried out by Mahmood et al (2002) was reported significant consumption of food material followed by gain in the larval weight through feeding "Farm yard manure and ammonia solution" treated mulberry leaves. There is relation among factors like the nutritional status of mulberry leaves and silkworm growth, silk yield and disease resistance Ravikumar (1988). According to Sengupta, et al (1972), nutrients like essential sugars, amino acids, proteins and vitamins are obligatory for normal growth of larval instars of silkworm. Javed and Gondal (2002) have reported higher growth and lower mortality of silkworm larvae fed with nitrogen and ascorbic acid supplemented mulberry leaves. Kanekatsu (1972; 1978); Eguchi and Iwamoto (1976); Abraham (1992) and Sumida, et al (1994) studied on midgut digestive enzymes of larval instars of silkworm, *Bombyx mori* (L). Kanekatsu, et al (1989) reported rationalization of some of midgut enzymes in larval instars of silkworm, *Bombyx mori* (L).

The *Syzygium cumini* (L) is a large evergreen tree, belong to family myrtaceae. It is a medicinal plant. Various parts of this plant are used in controlling the diabetes like diseases. The fruits and the seeds of *Syzygium* are used in folk medicine. The seeds of *syzygium* are excellent source of glycosides. The flavonol glycosides have been isolated from the roots of this plant. In one of the earlier studies in author's

laboratory, the glycosides are reported for the fortification of digestion in fifth instar larvae of silkworm, *Bombyx mori* (L). The seed powder of *Syzygium cumini* (L) is reported for contents of glycoside ( 5, 7-dihydroxy-6, 2 dimethoxyisoflavone-7-O-alpha-L-rhamnoside ) in earlier studies in laboratory of present attempt holders. The aim of present attempt is to analyze the effect of feeding the leaves of mulberry, *Morus alba* (L) aqueous solution of seed powder of *Syzygium cumini* (L) on the velocity of biochemical reactions catalyzed by midgut protease and midgut amylase in the fifth instar larvae of silkworm, *Bombyx mori* (L).

## II. MATERIAL & METHODS

The work was divided into the steps like: Silkworm Rearing; *Syzygium* solution Preparation; Grouping the Fifth Instar Larvae; Treating the mulberry leaves and feeding the larvae; Protein Bioassay and Statistical analysis.

**(A). Silkworm Rearing:** The egg cards or disease free layings (DFL) of bivoltine, crossbreed race: [(CSR6 x CSR26)] x [CSR2 x CSR27] of silkworm, *Bombyx mori* (L) were procured through the sericulture unit of Agriculture Development Trust, Malegaon. Black boxing was followed for incubation. The early age larvae (First and Second instared larvae) (Chawki) and late age larvae (Third; Fourth and Fifth instared larvae) were reared in the laboratory of "Dr. APIS" through the methods prescribed by Krishnaswami, et al (1978) & explained in earlier attempts by Khyade (2004); Vitthalrao & Kulkarni (2011); Desai, et al, (2011) Shubhangi Pawar, et al (2017); Ramprakash Verma, et al (2018); Pranita Rajendra Vare, et al (2018); Manisha Mahendra Nalwade, et al (2018); Seema K. Dongare, et al (2018) and the others. The larvae were fed with fresh and appropriate quality leaves of mulberry, *Morus alba* (L) procured from sericulture unit at Malegaon Sheti Farm of Agricultural Development Trust Baramati, Shardanagar, (Malegaon Khurd). The schedule of feeding prescribed by Sharad G. Jagtap

(2014) was followed for both early age larvae (First and Second instared larvae) (Chawki) and late age larvae (Third; Fourth and Fifth instared larvae). The fifth instared larvae were preferred for the analysis of effect of treating the mulberry leaves with aqueous solution of seeds of *Syzigium cumini* (L) and them for total protein contents.

**(B). Syzigium solution Preparation:** The ripen fruits of *Syzigium cumini* (L) were collected from Malegaon Sheti Farm of Agricultural Development Trust Baramati, Shardanagar, (Malegaon Khurd). They were identified and confirmed for species through the Botanical Survey of India, Pune. Seeds were separated and allowed for shade drying. It was followed by preparation of seed powder through the use of domestic mixture. Known quantity of this powder was kept for maceration in distilled water for twenty four hours. Macerated content was allowed for filtration through muslin cloth. Volume of filtrate and weight of residue were accounted for knowing the strength of seed powder in the solution. The filtrate was further utilized for preparation of aqueous solution known strength. Four different concentrations of solution were prepared, which include: 10 ppm; 20 ppm; 40 ppm and 50 ppm.

**(C). Grouping the Fifth Instar Larvae:** Soon after the fourth moult, the the fifth instared larvae were divided into six groups, each with hundred individuals. The groups include: Untreated Control; Water treated Control and four treated groups. The four treated groups include: 10 ppm; 20 ppm; 40 ppm and 50 ppm. 400 ml of aqueous solution of seed powder was used to treat 100 grams of fresh mulberry leaves. The treatment was carried out for half an hour before feeding. The treated mulberry leaves were drained off completely & then fed to the fifth instar larvae of silkworm, *Bombyx mori*(L) in respective groups. Feeding treated mulberry was carried out for the first four days of fifth instars.

**(D). Treating the mulberry leaves and feeding the larvae:** Mulberry leaf treatment was carried half an hour before each feeding. 2000 ml of aqueous solution of seed powder of each strength was used to treat 500 grams of fresh mulberry leaves for feeding the group of hundred larvae for each time. Fresh leaves of mulberry, *Morus alba* (L) were weighed. The known volume of solution of each strength was taken in separate glass jar. Known quantity of mulberry leaves was kept immersed separately in aqueous solution of each strength. The treatment was carried out for half an hour before feeding. The treated mulberry leaves were drained off completely & then fed to the fifth instar larvae of silkworm, *Bombyx mori* (L) in respective groups. Four feedings were followed ( 5.00 a.m.; 11.00 a.m. ; 5.00 p.m. ; 11.00 p.m.). Five hundred grams leaves of mulberry, *Morus alba* (L) were used for feeding the group of hundred larvae for each time. The feeding treated mulberry was carried out for the first four days of fifth instars. The larvae fed with untreated mulberry leaves and water treated mulberry leaves were also maintained.

**(E). Bioassay of midgut Soluble Proteins; midgut protease and amylase:** The bioassay of midgut Soluble Proteins; midgut protease and amylase was carried out on fifth day of fifth instar. Twenty larvae from each group were selected randomly. Weight of individual larva was recorded. They were anaesthetized with chloroform soaked cotton pads. Individual larva was dissected open from dorsal side. The entire alimentary canal was separated from individual larva. The alimentary canal was flushed with ice cold saline so as to remove the debris of mulberry leaf & washed with ice cold saline. The alimentary canal was blotted & weighed accurately on electronic balance. The mid gut tissue was fragmented & then homogenized in chilled saline. Homogenate was centrifuged at 400C for 15 min. at 10000 rpm. The supernatant was equalized to the volume, aliquots of which contain 10 mg per ml & used as assay sample. Half the volume of assay sample was utilized for bioassay of soluble

proteins & another half for mid gut enzymes (protease & amylase).

Bioassay of soluble proteins was carried out through the methods of Lowery, et al, (1951). For each assay sample ( of each group), bioassay was carried in the triplicate set. One ml of assay sample was added in each test tube. The blank test tube was also prepared simultaneously, in which the assay sample was replaced with distilled water. Addition of 5 ml Lowery's "C" solution was made in each test tube, mixed well & kept for 15 minutes for the purpose to form the copper-protein complex. After fifteen minutes; 0.5 ml Folin's phenol reagent was added in each test tube & mixed well. The content in each test tube was allowed to develop colour. Then the optical density of content of each test tube was recorded at 660 nm on spectrophotometer. The concentration of soluble proteins of each assay sample was calculated through the reference of optical density assay sample & standard proteins (BSA) (the plot of optical density against concentration of BSA).

The activity of mid gut protease was carried out according to the method of Brik, et al, (1962) with modifications suggested by Isshaya, et al, (1971) & outlined by Chougale (1992) & Khyade (2004). The mid gut protease activity was determined in triplicate set along with the blank. The mixture of incubation consisted of substrate ( one ml of ten percent casein solution) ; source of enzyme (0.5 ml assay sample) & 0.5 ml of 0.2M Trisbuffer (pH= 8.4). For the blank, assay sample was replaced by distilled water. The incubation was carried out in water bath at 300C for 20 minutes with constant shaking. Addition of 6 ml of 2 percent trichloroacetic acid was made. The content was centrifuged at 8000 rpm for 15 minutes. The supernatant was used to read the optical density at 280 nm on spectrophotometer. Amount of tyrosine liberated from the casein due to action of mid gut protease was calculated through the use of optical density readings for assay sample; tyrosine ( from standard graph) & predetermined soluble protein

contents of each assay sample. The activity of mid gut protease was expressed in terms of specific activity: microgram tyrosine liberated per mg protein per minute.

The activity of mid gut amylase was determined according to the methods of Bernfeld (1955); explained by Ishaaya & Swirski (1970), with modifications suggested by Gaikwad (1998) & outlined by Khyade (2004) & Desai, et al, (2011). For the purpose to determine the activity of mid gut amylase, 20 larvae were selected randomly & processed for assay sample preparation as described for soluble proteins. Mid gut amylase was determined in triplicate set along with blank. The incubation mixture consisted of one ml of one percent starch solution ( as substrate), phosphate buffer (pH=9.2) & 0.5 ml of assay sample. For the blank, assay sample was replaced by distilled water. The process of incubation was carried out in water bath at 300C for 20 minutes. After incubation the termination of activity of enzyme was made by addition of 2 ml DNSA & 2 ml distilled water. The contents were heated in boiling water bath exactly for five minutes, cooled immediately & the optical density of content was read at 540 nm on spectrophotometer.

For the purpose to calculate the mid gut amylase activity; the optical density readings for each assay sample; standard solution of maltase ( from graph) and soluble proteins were utilized. The enzyme activity was expressed in specific activity: micrograms of maltose liberated per mg protein per minute.

**(F). Statistical analysis:** Consistency in the results is qualitative parameter in research studies. Therefore, the whole experimentation in the present study was repeated for thrice. The data of all the three attempts was collected and subjected for statistical analysis. The statistical parameters for analysis considered in the study include mean, standard deviation, percent change & significance through student t – test introduced by William Sealy Gosset (a chemist

working for the Guinness brewery in Dublin, Ireland. ([https://en.wikipedia.org/wiki/Student%27s\\_t-test](https://en.wikipedia.org/wiki/Student%27s_t-test) )  
 "Student" was his pen name) and explained by Norman & Baily (1955).

**Table 1.** The Activity of Mid Gut Protease and Mid Gut Amylase in the Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]) Fed With the Leaves of Mulberry, *Morus alba* (L) (M-5: variety) Treated With Aqueous Solution of Seed Powder of *Syzigium cumuni* (L).

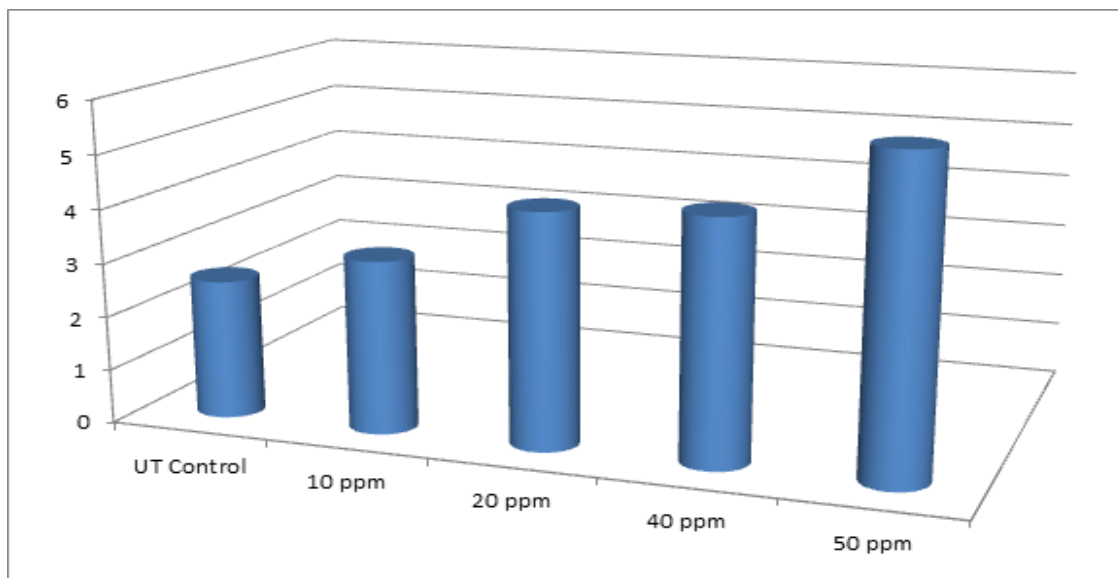
Mid Gut Enzymes Group	Protease	Amylase
Untreated Control	02.593 (±0.274) 00.000	5.547 (±0.816) 00.000
10 ppm	3.217* (±0.695) 24.064	6.864* (±1.213) 23.742
20 ppm	4.339** (±1.107) 67.335	10.148** (±2.321) 82.945
40 ppm	4.476*** (±1.786) 72.618	10.319*** (±3.312) 86.028
50 ppm	5.793*** (±2.011) 123.41	10.483*** (±3.786) 88.985

- Each figure is the mean & three replications.
- Figure in parenthesis with ± sign is the standard deviation.
- Figure below parenthesis is percent change.
- \* : P<0.05
- \*\* : P<0.01
- \*\*\* : P<0.001

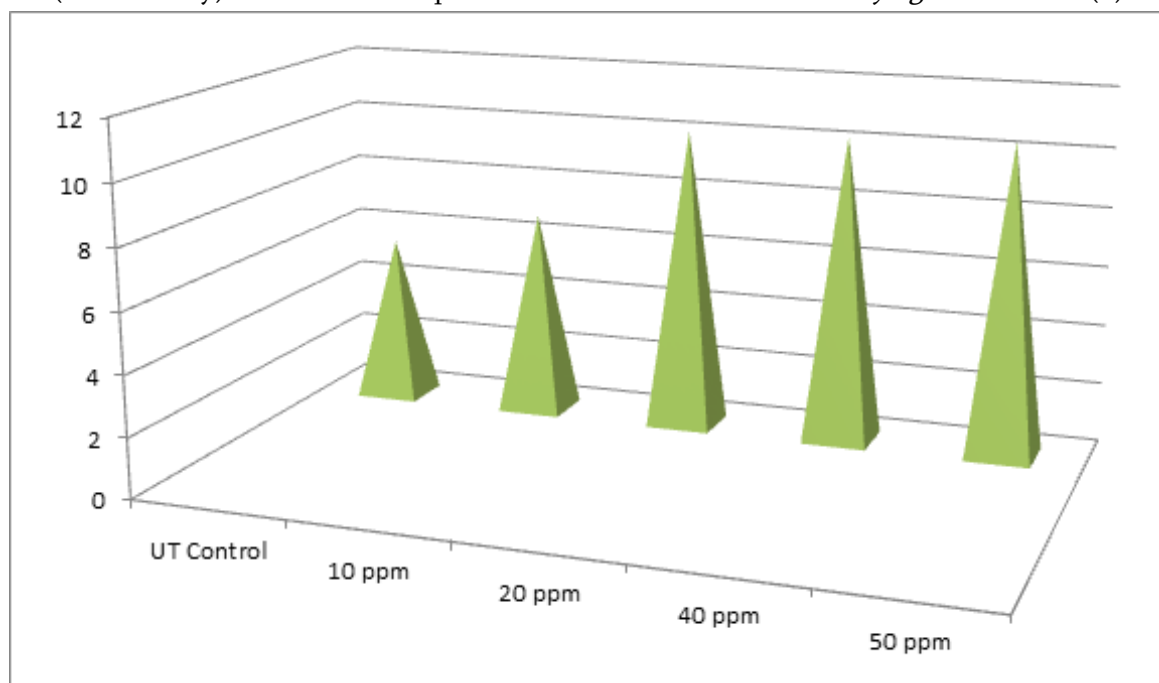
### III. RESULTS & DISCUSSION

The results on the effect of feeding the leaves of mulberry, *Morus alba* (L) aqueous solution of seed powder of *Syzigium cumuni* (L) on the velocity of biochemical reactions catalyzed by midgut protease and midgut amylase in the fifth instar larvae of silkworm, *Bombyx mori* (L). are summarized in table 1 & presented in Fig. 1 and 2. Treating the mulberry leaves with various concentrations of aqueous solution of *Syzigium* seed powder & feeding them to the fifth instar larvae of silkworm, *Bombyx mori* (L) for four

days was found variously reflected in the levels of activity of enzymes (protease & amylase) in the mid gut tissue homogenate.



**Figure 1.** The Activity of Mid Gut Protease in the Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]) Fed With the Leaves of Mulberry, *Morus alba* (L) (M-5: variety) Treated With Aqueous Solution of Seed Powder of *Syzigium cumuni* (L).



**Figure 2.** The Activity of Mid Gut Amylase in the Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]) Fed With the Leaves of Mulberry, *Morus alba* (L) (M-5: variety) Treated With Aqueous Solution of Seed Powder of *Syzigium cumuni* (L).

The velocity of biochemical reaction catalyzed by mid gut protease and midgut amylase in larvae fed with untreated mulberry leaves was found measured 02.593 units and 5.547 units respectively. The midgut protease activity in larvae fed with mulberry leaves treated with various concentrations (10.0 ppm; 20.0

ppm; 40.0 ppm & 50.0 ppm) of the aqueous solution of seed powder of *Syzigium cumuni* (L) was found measured 3.217; 4.339; 4.476 and 5.793 units respectively. The midgut amylase activity in larvae fed with mulberry leaves treated with various concentrations (10.0 ppm; 20.0 ppm; 40.0 ppm & 50.0

ppm) of the aqueous solution of seed powder of *Syzigium cumini* (L) was found measured 6.864; 10.148; 10.319 and 10.483 units respectively. Percent increase in the midgut protease activity through *Syzigium* treatment in present attempt was ranging from 24.064 to 123.41 (table – 1). Percent increase in the midgut amylase activity through *Syzigium* treatment in present attempt was ranging from 23.742 to 88.985 (table – 1). Significant improvement in the activities of midgut protease and amylase in the larval instars of silkworm, *Bombyx mori* (L) (Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]) fed with the leaves of mulberry, *Morus alba* (L) (M-5: variety) treated With aqueous solution of seed powder of *Syzigium cumini* (L) may be explained away as due to enhanced break down of contents of mulberry leaves. Some of the herbal powders contain insect juvenoids (like eugenol) are known to increase the capability of consumption & utilization of food by larval instars of insects like silkworm. The contents of seeds of *Syzigium cumini* (L) may have had such capabilities. This may be responsible for improve appetite & digestion.

According to Sen (1988), the plant derived compounds, in phytophagous insects, mimic the action of natural juvenile hormone, which enhance the synthesis of poly (A) RNA for major silk protein. Most significant response for *Syzigium* treatment in the study seems to be the levels of mid gut protease & mid gut amylase. The enzymes belongs to soluble proteins. The soluble proteins contribute in the tissue metabolism through enzymes. According to Applebaum (1985), continuous feeding in insects get reflect into advancement of production of mid gut enzymes, which improve the enzyme efficiencies. Most significant improvement in the protease activity in the treated group of study may be concerned with contents of specific plants. Individual plant extractive treatment may screen out the plant responsible for improved protease activity. Likewise the amylase enhancing herbal constituents of herbal formulations should be screened.

Feeding treated mulberry leaves for first four days possibly availing the herbal nutrients, which affect digestibility of larvae & may contribute phytojuvenoids or other compounds of growth & development. The study should be extended for screening juvenoid activity of *Syzigium* seed powder.

#### IV. ACKNOWLEDGEMENT

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#### V. REFERENCES

- [1]. Applebaum, S.W. (1985). Biochemistry of digestion. In: Comprehensive insect physiology (Ed. Kerkut, G.A. and Gilbert (I), 4:297-307. Pergamon press New York.
- [2]. Bernfeld, P. (1955). Amylase, a and b. In: Methods of Enzymology, Vol.I (Ed. Clockwik & Kalpin). Academic Press, New York.
- [3]. Bowers, W.S.; Fales, V.M.; Thompson, M.J. and Uebel,B.(1966). Juvenile & gonadotropic activity of 10,11 epoxyfranesoic acid methyl ester. Life Science. 4: 2323-2331.
- [4]. Brik, Y.; Harpaz,J. ; Ishaya and Bhondi, A.(1962), Studies on proteolytic activity of beetle, *Tenebrio molitor* (L). J. Insect Physiol. 8: 417-429.
- [5]. Chougale, A.C.(1992). Influence of magnetic energy on silkworm, *Bombyx mori* (L). Ph.D. Thesis, Shivaji University, Kolhapur.
- [6]. Desai, V.A.; Pawar, V.V. & Sawant, R.T.(2011). Influence of herbal drug: kho-go on the fifth instar larvae of silkworm, *Bombyx mori* (L).Dissertation in the partial fulfillment of M.Sc. (Microbiology), Shardabai Pawar Mahila College, Shardanagar (Baramati), (Pune University, Pune).



- [7]. Gaikwad, A.R.(1998). Biology of some dung beetles of South Western Maharashtra. Ph.D. thesis, Shivaji University, Kolhapur.
- [8]. Ghantaloo, U.S. (2007). Influence of digoxin on silkworm, *Bombyx mori* (L). M.Phil. Thesis, Alagappa University, Karaikudi (Tamil Nadu), India.
- [9]. Horie, Y.; Tanaka, M. and Ito, T. (1963). Proteolytic enzyme of digestive juice of mid gut in silkworm, *Bombyx mori* (L). *J. Sericult. Sci. Japan*, 32: 8-15.
- [10]. Horie, Y.(1961). Physiological studies on the alimentary canal of silkworm, *Bombyx mori* (L). III. Absorption & utilization of carbohydrates. *Bull. Sericult. Exp. Sta. Japan*, 16: 287-309.
- [11]. Ishaya, I. and Swirski, E. (1976). Trehalase, invertase & amylase activities in the larvae of Egyptian cotton worm, *Spodoptera littoralis* (L). *J. Insect Physiol.* 17: 945-953.
- [12]. Ito, H. (1960). Effect of sugars on feeding the larvae of silkworm, *Bombyx mori* (L) *J. Insect. Physiol.* Vol.5: 95-107.
- [13]. Ito, T.(1961). Nutrition of silkworm, *Bombyx mori* (L). *Proc. Jpn. Acad. Sci.* 43:57-61.
- [14]. Ito, T; Kwashima, K.; Nakhara, M.; Nakanshi, K. & Terahara, A.(1964). Metabolism in the mid gut of silkworm, *Bombyx mori* (L). *Insect Physiol.* Vol.10: 225-228.
- [15]. Ishaaya, I.; Moore, I and Joseph, B.(1971). Protease & amylase activity in the larvae of Egyptian cotton worm, *Spodoptera littoralis* (L). *J. Insect physiol.* 17: 945-953.
- [16]. Jagtap, S.G.(2007). Effect of plant juvenoids on consumption & utilization of mulberry leaves by silkworm, *Bombyx mori* (L). M.Phil. Thesis, Alagappa University, Karaikudi, Tamil Nadu, (India).
- [17]. Khyade, V.B.(2004). Influence of juvenoids on silkworm, *Bombyx mori* (L). Ph.D. thesis, Shivaji University, Kolhapur.
- [18]. Krishnaswami, S.; Narasimhna, M.N.; Suryanarayana, S.K. and Kumararaj, S.(1978). *Sericulture Manual-II Silkworm Rearing*: FAO. United Nations Rome.
- [19]. Lowery, O.H.; Rosenbrough, N.J.; Far, A.L. and Randall, R.J.(1951). Protein measurement with folin phenol reagent. *J. Biol. Chem.* 193: 265-275.
- [20]. Murali, K.(1992). Effect of leaf carbohydrate reserves on the growth & excretory pattern of silkworm, *Bombyx mori* (L). M.Phil. Dissertation, Sri. Venkateshwara University, Tirupati (India).
- [21]. Murugan, K. and George, A. (Sr.)(1992). Feedings & nutritional influence on growth & reproduction of *Daphnia* near (L). *Insect Physiol.* 38: 961-969.
- [22]. Nayar, J.K. & Frankel, G.(1962). *Journal of Insect Physiology*. Volume-8, page-505.
- [23]. Norman, T.J. & Baily (1955). *Statistical methods in Biology*.
- [24]. Slansky, F. & Scriber, J.M. (1985). Food consumption & Utilization. In: *comprehensive Insect physiology, Biochemistry & pharmacology*. (Eds. Kerkut, G.A. and Gilbert, L.I.) 4, Pergamon Press, Oxford, Page: 639.
- [25]. Subramanyam Reddy, C.(1992). Studies on distribution of digestive enzymes in the digestive tract of silkworm, *Bombyx mori* (L). M.Phil. Dissertation, Sri. Venkateshwara University, Tirupati (India).
- [26]. Vitthalrao Khyade and Jeevan P. Sarawade (2009) protein profiles in the fifth instar larvae of silkworm, *Bombyx mori* (L)(PM x CSR2) fed with digoxin treated mulberry leaves. *The Bioscan*(1): 41-44.
- [27]. Vitthalrao B. Khyade & Jyoti Kulkarni (2011). Effect of digoxin treated mulberry leaves on protein profiles in fifth instar larvae of silkworm, *Bombyx mori* (L) (PM x CSR2). *Res. J. Chem. Sci.* Vol.1 (1): 2-7 ([www.isca.in](http://www.isca.in))
- [28]. Yamafuji, I. and Yonezawa(1935). Lipases in silkworm, *Bombyx mori* (L). *Insect. Biochem.* 1: 102-112.

- [29]. Kellner O, Kakizaki S, Matsuoka M, Yoshu T. XXIV. On the physiology of the silk workm. By Alexander pringle jameson and william ringrose gelston atkins. Landw. Versuchs-stationen. 1887; 33: 38.
- [30]. Mahmood R, Jan MT, Khan MI, Effect of nitrogen (farmyard manure + urea) treated mulberry trees on the larval development and cocoon weight of silkworm, *Bombyx mori* L. *Asian J plant Sci.* 2002; 2 (1): 93-94.
- [31]. Ravikumar C. Western ghat as a bivoltine region prospects, challenges and strategies for its development. *Indian Silk.* 1988; 26(9): 39-54.
- [32]. Sengupta K, Singh B.D, Mustafij C. Nutrition of silkworm. *Bombyx mori* L.I. Studies on the enrichment of mulberry leaf with various sugars, proteins, aminoacids and vitamins for vigorous growth of the worm and increased cocoon crop production. *Indian J Sci.* 1972; 11:11-27.
- [33]. Javed H, Gondal MH. Effect of food supplementation by N and Ascorbic Acid on larval mortality of silkworm (*Bombyx mori* L). *Asian journal of plant science.* 2002; 1(5): 556-557.
- [34]. Kanekatsu R. Amylase in the digestive juice of silkworm larvae, *Bombyx mori*. *J Seric Sci.* 1972; 41: 445-451.
- [35]. Kanekatsu R. Studies on further properties for an alkaline amylase in the digestive juice of silkworm, *Bombyx mori*. *J Fac Text Sci Technol.* 1978; 76: 1-21.
- [36]. Eguchi M, Iwamoto A. Alkaline protease in the midgut tissue and digestive fluid of silkworm, *Bombyx mori* L. *Insect Biochem.* 1976; 6: 491-496.
- [37]. Sumida M, Yuan X L, Matsubara F. Sucrase activity and its kinetic properties in peritrophic membrans, and in membrane-bound and soluble fractions of midgut in silkworm, *Bombyx mori* L *Comp Biochem Physiol. A* 1994; 108: 255-264.
- [38]. Abraham E G, Nagaraju J, Datta RK. Chemical studies of amylases in the silkworm, *Bombyx mori* L Comparative analysis in diapause and nondiapause strains. *Insect Biochem Mol Biol.* 1992; 22: 867-873.
- [39]. Kanekatsu R, Ichimura H, Hori M. Distribution and developmental changes in midgut sucrase activity of the silkworm, *Bombyx mori*. *J Seri Sci Japan.* 1989; 58: 517-523