

Phytochemical Studies and Antidiabetic Activities of Newbouldia laevis (P. Beauv) Ethanolic Leaves Extracts in Alloxan-Induced Diabetic Rats

Grace D. O. Eneh¹, Okon G. Okon^{2*}, Joseph E. Okon³ and Nsikak B. Essien¹

¹Department of Science Technology, Akwa Ibom State Polytechnic, Ikot Osurua, Akwa Ibom State, Nigeria
 ²Department of Biological Sciences, Ritman University, Ikot Ekpene, Akwa Ibom State, Nigeria
 ³Department of Biological Sciences, Akwa Ibom State University, Ikot Akpaden, Akwa Ibom State, Nigeria

ABSTRACT

The phytochemical properties of the ethanolic leaves extract of Newbouldia laevis and its antidiabetic properties on alloxan-induced diabetic albino rats were evaluated. The qualitative phytochemical screening revealed the presence of alkaloids, tannins, saponins, terpenes, flavonoids, cardiac glycosides and anthraquinones in Newbouldia laevis leaf extract. Quantitative phytochemical screening of Newbouldia laevis revealed the presence of flavonoids ($12.22 \pm 1.67 \text{ mg/kg}$), alkaloids ($10.71 \pm 1.75 \text{ mg/kg}$), saponins (7.01 ± 0.61 mg/kg), tannins (6.74 ± 0.36 mg/kg), cardiac glycosides (4.11 ± 0.46 mg/kg), anthraquinones (1.42 ± 0.24 mg/kg) and terpenes 1.16 ± 0.24 mg/kg. The toxicity test of the extracts produced various degree of toxicity ranging from writhing, decreased respiration, convulsion to mortality. The intensities of these effects were proportional to the dose administered. The LD50 for Newbouldia laevis was 447.21 mg/kg. The results of the influence of ethanolic leaves of *Newbouldia laevis* on alloxan-induced diabetic rats showed significant (p<0.05) reduction in the blood glucose level (BGL) as time of treatments increases. The leaves extracts of Newbouldia laevis significantly (p<0.05) decreased the blood glucose level of diabetic rats on day 14 compared to diabetic untreated (control) rats. The BGL of diabetic rats treated with oral hypoglycemic agent alone were significantly (p<0.05) different from control group on day 14. The presence of these biologically active compounds suggest that the plant could serve as potential sources of drugs and their secondary metabolites could exert some biological activities when taken by animals. The extracts of Newbouldia laevis also produced a similar reduction in blood sugar level as the standard hypoglycemic drug (Glibenclamide). This may be due to its ability to enhance insulin secretion like the standard drug. This positive result confirms why herbalists use this leaf for the management of diabetes.

Keywords : Albino Rats, Alloxan, Antidiabetic, Diabetes, Glibenclamide, Newbouldia laevis, Phytochemicals

I. INTRODUCTION

Diabetes mellitus (DM) is one of the most common non-communicable diseases globally [1]. It is either the fourth or fifth leading cause of death in developed and developing countries of the world [2]. Symptoms of DM include classical hyperglycaemia, polyuria, polydypsia, weight loss, sometimes with polyphagia, and blurred vision. Diabetes mellitus is classified into Type-1 or Type-2 depending on the aetiology [3].

Plant drugs are frequently considered to be less toxic and free from side effects than synthetic chemicals [4].

Before the advent of insulin and chemical drugs for the treatment of diabetes as with other ailments, plant-based medications have been used as traditional remedies for the treatment of many diseases including diabetes mellitus [5, 6, 3]. More than 400 plants are being used in different forms for their hypoglycaemic effects in treating diabetes [7, 3]. In Nigeria, traditional medicine occupies a unique position in health care delivery, especially among the rural populace. However, the activities of herbalists are surrounded with a lot of secrecy and lack of scientific procedure, hence the need to standardize the practice of traditional medicine [8, 3].

The use of medicinal plants for treatment and management of diseases is gaining prominence worldwide especially in the developing countries where 80 % of the population still depends on traditional healing methods [9, 10]. This surge in the use of herbal medicines is probably due to the perceived failure of some synthetic drugs in the treatment of some diseases, the side effects associated with most drugs and the incidence of drug resistance especially among the antibiotics family [11]. In recent times, quite a number of some plants have been used as herbal medicines due to the presence of phytochemicals in them [12]. Many of these discovered phytochemicals seem to fight diseases and lower the rates at which they occur [13].

N. laevis is a medium size angiosperm which belongs to the *Bignoniaceae* family. It is a fast-growing evergreen shrub or small tree. It only reaches a height of 3 - 8 metres in the west of its range, but can attain a height of up to 20 metres in the east. It has many stem forming clumps of gnarled branches [14].

The tree, and especially the bark, is widely used in traditional medicine in Africa. *Newbouldia laevis* is one of such plants that its leaves are used in Southeastern Nigeria to hasten parturition and to expel the placenta after delivery. Agents that stimulate uterine contraction are classified as oxytocics and are employed clinically for the induction and support of labour as well as in the management of the third stage of labour [15].

The bark has analgesic and stomachic properties. A decoction is used in the treatment of coughs, diarrhea and dysentery, while it is also given to children for the treatment of epilepsy and convulsions. A decoction of the bark, combined with chillies, is used in the treatment of chest pains [15]. The dried bark and young twigs, pounded up with spices such as *Xylopiasp*, are given in decoction or infusion to treat such complaints as uterine colic, dysmenorrhoea, etc [15]. Thus, the aim of this study was to assess the phytochemical properties and antidiabetic effect of the *Newbouldia laevis* ethanolic leaf extract on albino rat.

II. METHODS AND MATERIAL

A. Study Area

This research was carried out in Uyo Local Government Area of Akwa Ibom State, Nigeria. Uyo is a city in South-South Nigeria found between latitude 5.02°N and longitude 7.92°E; it has an average temperature of 25.1-27.8°C and an annual rainfall range of 33-37.8mm with the land mass of 115 km² and the population of 1,400 thousands persons/km².

B. Plants Collection and Authentication

The leaves of *Newbouldia laevis* were collected from plants growing in the University of Uyo farm, University of Uyo, Akwa Ibom State. The plant samples used for this research work were authenticated by a Plant Taxonomist in the Department of Botany and Ecological Studies, University of Uyo, Uyo, Nigeria.

C. Collection and Maintenance of Experimental Animals

Male and female albino rats weighing 150 – 250g were obtained from the animal house of the

Department of Biochemistry, University of Uyo and randomly assigned to five groups of five rats in each group; Two groups served as a positive control and a negative control, while the other three groups served as test groups, which were used for application of *Newbouldia laevis* leaves extract.

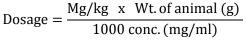
D. Preparation of Plant Extracts

The fresh leaves of *Newbouldia laevis* were air dried for 7 days and ground into powdered form, 400g of each plant samples were extracted using 6000ml of 70% ethanol and shaken intermittently for 72 hours. It was filtered and the filtrate concentrated (dried) *invacuo* at 40°C in a water bath. The extract was weighed and stored in 150ml beaker, labeled and covered with foil paper and preserved in the refrigerator at 4°C for use in qualitative and quantitative phytochemical screening, acute toxicity test, and anti-diabetic tests.

E. Preparation of Extract and Fraction Solutions

The extract and fraction solutions were prepared by dissolving about 1.5g (using high performance profile design compact weighing balance: CS200) of the extract and fractions in 10 ml of distilled water, to give an effective concentration.

The formula:



was used to calculate the volume of the extract/fractions solution to be administered to each animal. The solutions were prepared fresh daily before administration [16].

F. Treatment Application and Experimental Design

Three groups out of five groups of rats were administered with ethanolic extract of Newbouldia laevis. Group one and two were used as control (positive and negative control). The positive control was orally given soluble fraction of standard drug, fed with normal feed and distilled water. Group two were fed with normal feed and distilled water. Group three were administered orally with 40% (LD) soluble fraction of ethanolic extract of Newbouldia laevis, normal feed and distilled water. Group four were fed orally with 60% (MD) soluble fraction of ethanolic extract of Newbouldia laevis, allowed free access to normal feed and distilled water. Group four were administered orally with 100% (HD) soluble fraction of ethanolic extract of Newbouldia laevis, allowed free access to normal feed and distilled water daily for 14 days (two weeks). The animals were anaesthetized and sacrificed with chloroform. Blood samples were collected with 5 ml syringe by cardiac puncture into ethylene diaminetetraacetic acid (EDTA) sequestrene bottles to prevent coagulation and used for determination of antidiabetic activities within 24 hours of sample collection.

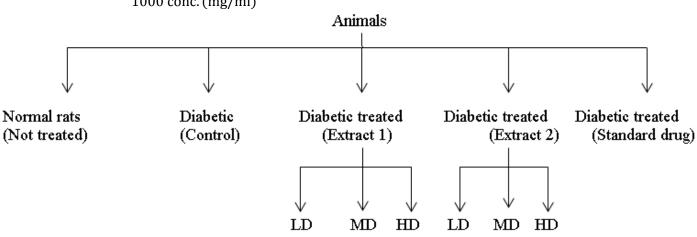


Figure 1. Treatment Application and Experimental Design

G. Determination of Median Lethal Dose (LD50)

Albino rats weighing 25 – 32g were kept in five cages (5 per cage) and handled according to standard guidelines for the use and care of laboratory animals. Food was withdrawn for 18 hours before the onset of the experiment [17]. The five groups of mice were administered with 4000mg/kg, 3000mg/kg, 2000mg/kg, 1000mg/kg and 500mg/kg of *Newbouldia laevis* extract. The groups were observed for mortality rate within 24 hours and the median lethal dose (LD₅₀) was calculated according to the methods of Lorke [18] with this formula:

$$LD_{50} = \sqrt{D_0 \times D_{100}}$$

 $\label{eq:where: D_0 = Maximum dose producing 0\%} Where: D_0 = Maximum dose producing 0\%$ mortality

 $D_{100} \ = \ Minimum \ dose \ producing \ 100\%$ mortality

H. Determination of Weight, Feed and Water intake of the Rats

Feed and water intakes were measured every day at the same hour during the experimental periods while the body weight of the animals was measured at zero day and every seven days for the period of 21 days using high performance profile design compact weighing balance (DW 1100) [19].

I. Determination of Blood Glucose

Blood samples were collected at 0, 30, 60 and 120 minutes post-treatment by a tail tip-snip cut. The plasma glucose concentration of the rats was determined with the aid of an electronic glucose meter (Accu-Chek Advantage) and glucose strips (Accu-Chek Advantage II). The mean antidiabetic response observed was recorded as mmol/L and used to determine the dose response effect of *N. laevis* [3].

J. Qualitative Phytochemical Screening

The methods of [20, 21, 22] were used for qualitative phytochemical screening of the leaf extracts. These included tests for saponins, tannins, flavonoids, anthraquinones, terpenes, phlobatannins alkaloids and cardiac glycosides. The experiment was carried out in the Department of Pharmacognosy and Natural Medicine, University of Uyo, Uyo.

K. Antidiabetic Analysis

The alloxan-induced diabetic albino rats were fasted and placed in 5 groups (groups 2 - 6) of 5 rats each and treated as follows: group 1(control rats) received normal saline orally for14 days; group 2 were untreated diabetic rats; groups 3, 4 and 5, which were all diabetic, received orally 100, 200, 400 mg/kg of the extract, respectively, for 14 days. The 6th group (diabetic) received 5 mg/kg of glibenclamide orally for 21 days. These doses were selected after an initial preliminary work done using various doses on diabetic rats.

L. Statistical Analysis

The statistical analysis was carried out to find the effect of *Newbouldia laevis* diet on antidiabetic parameters in rats. One way analysis of variance was adopted for comparison and the results are expressed as standard error of mean (S.E.M.). The significant difference between means was determined using one way ANOVA at significance level of p < 0.05 according to the methods of [23].

III. RESULTS AND DISCUSSION

Qualitative phytochemical screening of ethanolic leaf extracts of *Newbouldia laevis* revealed the presence of some bioactive constituents, as summarized in Table 1. Ethanolic leaf extracts of *Newbouldia laevis* showed the presence of alkaloids, tannins, saponins, terpenes, flavonoids, anthraquinones and cardiac glycosides. Quantitative phytochemical screening of *N. laevis* recorded that flavonoids had the highest value $(12.22 \pm 1.67 \text{ mg/kg})$, followed by alkaloids with $(10.71 \pm 1.75 \text{ mg/kg})$, saponins had $(7.01 \pm 0.61 \text{ mg/kg})$

mg/kg), tannins (6.74 \pm 0.36 mg/kg), cardiac glycosides (4.11 \pm 0.46 mg/kg), anthraquinones (1.42 \pm 0.24 mg/kg) while terpenes recorded the lowest value with 1.16 \pm 0.24 mg/kg (Table 2).

Constituents	Test	Observations	N. laevis		
Alkaloids	Dragendoff's test	Formation of red	+++		
		precipitate			
Tannins	Ferric Chloride	A blue-green	+++		
	Test	precipitate			
Saponins	Frothing Test	Appearance of froth.	+++		
	Forming Test	Formation of 1 layer of	+++		
		form			
Terpenes	Libermanns	Formation of brown	+		
	Burchard's test	ring at the junction			
Flavonoids	Shinoda's Test	a red colour indicated	++		
		the presence of			
		flavonoids			
Phlobatannins		No visible colour	ND		
Anthraquinone	Borntrager's test	the presence of a pink	ND		
		in the ammoniacal			
		(lower) phase			
	Sulphuric acid	a violet coloration in	ND		
	test	the ammonia layer			
		(Lower phase)			
Cardiac	Salkowski test	a reddish-brown colour	+		
glycocides		at the interphase			
	Keller-kellani's	brown ring obtained at	+		
	test	the interface			

Table 1. Qualitative phytochemical screening of Newbouldia laevis

+ = Trace, ++ = Moderate, +++ = Abundant, ND = Not detected

Table 2.	Quantitative	phytochemical	screening of N.
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	laevis
Phytochemicals	<i>N. laevis</i> (mg/kg)
Alkaloids	10.71 ± 1.75
Flavonoids	12.22 ± 1.67
Tannins	6.74 ± 0.36
Saponins	7.01 ± 0.61
Cardiac glycosides	4.11 ± 0.46

Terpenes	1.16 ± 0.04
Anthraquinones	1.42 ± 0.24

Data is presented as mean \pm standard error (\pm SEM) of triplicate values

The importance of alkaloids, saponins and tannins in various antibiotics used in treating common pathogenic ailments have been reported [24]. The medicinal value of plants lies in their inherent chemical substances which have a definite physiological action on the human body [25]. Different phytochemicals have been found to possess a wide range of activities which may help in protection and prevention against chronic diseases. For example alkaloids are known to protect against diabetes, liver infection and malaria. Saponins protect the body against hypercholesterolemia and possess antibiotic properties while terpenes show analgesic properties [26]. Esenowo et al. [27] reported that tannins, flovanoids and cardiac glycosides in Peristrophe bicalculata have the potency to promote haemopoictic indices and restore loss of blood during excessive bleeding. Similar reports were obtained for Morinde *citrotolia* and *Digitalis pupurea* by Farine *et al.* [28]; Trease and Evans [20]. They reported the presence of cardiac glycosides in these plants and stated that they can be used in the treatment of diseases associated with the heart. These plants are currently used by herbalists to treat tumour [29]. Flavonoids containing plants have been used as

diuretic, laxative, emollient and poultice [30]. Therefore the use of these plants rich in flavonoids and other chemical substances in traditional medicine lends credence to the medicinal potentials of these plants. Terpenes in modern clinical studies have supported the role of plants as anti-inflammatory and analgesic agent [31].

To access the toxicity of *N. laevis*, the rats were treated intraperitoneally with a single dose of 0.09 - 0.64 mg/kg of *N. laevis* soluble fraction after being starved for 18 hours. Intraperitoneal route was chosen because of its sensitivity and rapid results. These ethanolic leaf extracts produced various degree of toxicity ranging from writhing, decreased respiration, excitation, decreased motor activity, convulsion to mortality (Table 3). The intensities of these effects were proportional to the dose administered. The intraperitoneal LD₅₀ for *N. laevis* it was 447.21 mg/kg.

Groups	Dose (mg/kg)	Average weight of	Number of mice	Percentage	
		mice (g)	per group	mortality	
Ι	100	21	5	0/3	
II	200	22	5	0/3	
III	300	22	5	0/3	
IV	400	23	5	0/3	
V	500	20	5	3/3	

 Table 3. Toxicity of N. laevis ethanolic leaf extract on albino mice

 $LD_{50} = \sqrt{AB}$

Where A = 400 (Maximum dose producing 0% mortality)

B = 500 (Minimum dose producing 100% mortality)

 $LD_{50} = \sqrt{400} \ x \ 500$

 $LD_{50} = 447.21 \text{ mg/kg}$

Table 4 summarizes the effect of ethanolic leaves extract of *N. laevis* on alloxan-induced diabetic rats. The results showed a significant (p=0.05) reduction in the blood glucose level (BGL) as the time of treatments increases. At 0 hour, the negative control (normal saline) recorded 332.0 ± 19.0 mg/dl, positive control (Glibenclamide) was 340.0 ± 19.9 mg/dl, low dose (308.0 ± 19.2 mg/dl), middle dose (316.0 ± 25.2) while high dose recorded 314.0 ± 7.42 mg/dl. While high dose recorded 314.0 ± 7.42 mg/dl. At 24 hours, the following values were recorded; negative

control (352.0 \pm 18.1 mg/dl), positive control (192.0 \pm 12.7 mg/dl), low dose (184.0 \pm 4.26 mg/dl), middle dose (122.0 \pm 3.06 mg/dl) and high dose (120.0 \pm 7.94 mg/dl). At the end of the treatments i.e. 14 days, the increase was persistent in the negative control (321.0 \pm 9.54 mg/dl), progressive reduction was recorded in the positive control (97.3 \pm 7.31 mg/dl), low dose was 139.0 \pm 4.91 mg/dl, middle

dose (95.3 \pm 4.63 mg/dl) while high dose recorded 89.7 \pm 2.33 mg/dl. Comparatively, the values recorded at 14 days of constant treatment in positive control, middle dose and high dose were within the same range (89.7 – 97.3 mg/dl) (Table 4).

Dosage	Fasting Blood glucose level at time interval (mg/dl)									
(mg/kg)	Initial	0hour	1hour	2hours	3hours	5hours	7hours	24hou	7days	14days
								rs		
Control	75±5.5	332±1	336±1	338±19.	340±19.	342±1	342±1	352±1	331±1	321±9.5
(-ve	7 ª	9.0ª	9.50ª	30 ª	2ª	8 .1ª	8.5ª	8 .1ª	4.6ª	4 ^a
control)										
Glib. 5	75.7±3	340±1	321±2	296±21.	275±22.	224±3	187±3	192±1	158±3	97.3±7.
(+ve	.67ª	9.9ª	1.10 ^b	10 ^b	8 ^b	6.2 ^ь	9.5 [⊾]	2.7 ^b	0.9 ^b	31 ^b
control)										
NL 44.72	76±5.1	308±1	294±1	277±20.	238±10.	220±1	197±1	184±4.	153±7.	139±4.9
(LD)	3 ª	9.2 ^b	9.70 ^c	30°	1°	7.3 [⊾]	7.1°	26°	31 ^b	1°
NL 89.44	81.7±6	316±2	300±2	245±8.1	206±5.6	165±4.	122±2.	122±3.	105±2.	95.3±4.
(MD)	.36ª	5.2°	5.20 ^d	1^{d}	9 ^d	1 8 °	03 ^d	06 ^d	6 ^c	63 ^b
NL 134	82.7±7	314±6.	293±7.	266±8.6	234±9.4	189±9.	144±9.	120±7.	99±2.0	89.7±2.
(HD)	.42ª	64°	88ac	5 ^e	9 °	94 ^d	39 ^e	94^{d}	8 ^c	33 ^b

Table 4. Anti-diabetic effects of ethanol leaves extract of *N. laevis* on alloxan induced diabetic

Values are expressed as Mean \pm SEM; Glib = Glibenclamide, HD = High dose, MD = Middle dose, LD = Low dose, NL = N. *laevis*, -ve = Negative, +ve = Positive. a-b Means with different superscripts along the same column are significantly different (p<0.05).

Induction of diabetes using alloxan has been described as a useful experimental model for studying the effect of hypoglycemic agents [32]. Alloxan and the product of its reduction, dialuric acid, establish a redox cycle with the formation of superoxide radicals. These radicals undergo dismutation to hydrogen peroxide with a simultaneous massive increase in cytosolic calcium concentration, resulting in the destruction of pancreatic β -cells and diabetes [32]. The untreated diabetic rats had a significantly higher fasting blood glucose level than normal rats that received treatment. This confirms induction of diabetes by

alloxan. The results showed a dose-dependent lowering of fasting blood glucose level in diabetic rats treated with different doses of the ethanol extracts of *N. laevis.* This dose-dependent effect compares well with glibenclamide, the reference drug used. Glibenclamide is a standard drug that is routinely used in the treatment of diabetes [33].

It is possible that the ethanolic leaf extracts of the test plants could have induced insulin secretion just like those treated with oral hypoglyceamic drug (glibenclamide). This positive result confirms why traditional medicine practitioners use the leaves of the plants in folk medicine in the treatment of

of insulin by diabetes. Hence the release glibenclamide produces а lowering of hyperglycemia. The extract of N. laevis also produced a similar reduction in blood sugar level and it is probable that it also enhanced insulin secretion but this needs to be investigated in future studies. The preliminary identification of alkaloids, glycosides and their co-existence with phenolic acids may explain at least in part some anti-diabetic properties of the extracts. The observed antihyperglycemic activity was in agreement with the findings of a study, which showed that ethanol leave extract of N. laevis decreased blood glucose level in diabetic rats [34]. However, the presence of flavonoids in the ethanol extracts may account for the observed hypoglycaemic effect since they have been found to stimulate the secretion of insulin [35]. Glibenclamide (an oral hypoglycemic agent) is known to act by enhancing exogenous insulin contribution which thus corrects deficiency in the endogenous insulin created by alloxan.

IV. CONCLUSION

The results from this study showed that ethanolic leaf extract of N. laevis contain a number of bioactive constituents such as; alkaloids, tannins, saponins, terpenes, flavonoids and cardiac glycosides. The presence of these biologically active compounds suggest that the plant could serve as potential sources of drugs and their secondary metabolites could exert some biological activities when taken by animals. The results also showed that of *N. laevis* ethanol leaves extract significantly reduced blood glucose of diabetic rats on day 14 compared to diabetic untreated (control) rats. The extracts of N. laevis therefore produced a similar reduction in blood sugar level as glibenclamide standard drug. This positive result may explain why some people use the plant for diabetes treatment.

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VI. REFERENCES

- Kaviarasan, S., Muniandy, S., Qvist, R., Chinna, K. and Ismail, I. S. (2010). Is F2isoprostane a biological marker for the early onset of Type 2 diabetes mellitus? *International Journal of Diabetes for Developing Countries*. 30(3): 165-170.
- [2] Amos, A. F., McCarty, D.J. and Zimmet, P. (1997). The rising global burden of diabetes and its complications: Estimates and projections to the year 2010. *Diabetic medicine*. 14: 1-85.
- [3] Ezeigbo, I. I. (2010). The Antidiabetic Activities of the Methanolic Leaf Extract of *Hymenocardi aacida* (Tul). Published MSc. dissertation. University of Nigeria, Nsukka. 8 – 22pp.
- [4] Momin, A. (1987). Role of indigenous medicine in primary health care In: Proceedings of 1stInternational seminar on Unani medicine, New Delhi, 54p.
- [5] Jayakumar, R. V. (2010). Herbal medicines for Type 2 diabetes: Guest editorial. *International Journal of Diabetes for Developing Countries*. 30 (3): 111-112.
- [6] Bhagwat, D.A, Killedar, S. G. and Adnaik, R.
 S. (2008). Anti-diabetic activity of leaf extract of *Tridax procumbens. International Journal of Green Pharmacy*, 2: 126-128.
- [7] Iwu, M. M., Okunji, C. O. and Ohiaeri, G. O. (1990). Hypoglycaemic activity of dioscoretine from tubers of *Dioscore adumetorum* in normal and alloxan diabetic rabbits. *Planta Medica*, 56: 264-267.
- [8] Madubuike, K. G. (2007). The antiinflammatory activities of the leaf

chloroform extract of *Palisota hirsute* (K. SCHUM). Unpublished MSc. dissertation. University of Nigeria, Nsukka.

- [9] Iwu, M. M. (1993). Handbook of African medicinal plants. Boca Raton, FL: CRC Press. 4-10pp.
- [10] De-Silva, T. (1997). Medicinal plants for forest conservation and health care: industrial utilization of medicinal plants in developing countries. Global Initiative for Traditional Systems (gifts) of Health. of Food and Journal Agricultural Organization of the United Nations, 11: 34-44.
- [11] Lewis, S. M., Bain, B. J. and Bates, I. (2001).
 Dacie and Lewis: Practical Haematology.
 Ninth Edition, London, New York: Elsevier, 9-40pp.
- [12] Riby, J. E. (2006). Herbal medicines and phytochemicals of indegenoues plants in California. *Berkely Molecular and Pharmarcology Journal*, 69(2): 430-439.
- [13] Harborne, J. B. (1973). *Phytochemical methods*, London: Chapman and Hall, 110-113pp.
- [14] Arbonnier, M. (2004). *Trees, shrubs and lianas of west african dry zones*. Germany: CIRAD Press Ltd. 243 248pp.
- [15] Bafor, E. and Sanni, U. (2009). Uterine contractile effects of the aqueous and ethanol leaf extracts of *Newbouldia laevis* (Bignoniaceae) *in vitro*. *Indian Journal of Pharmaceutical Science*, 71(2): 124–127.
- [16] Okpuzor, J., Ogbunugafor, H. A. and Kareem, G. K. (2009).Hepatic and hematologic effects of fractions of *Globimetula braunii* in normal albino rats. *EXCLI Journal.* 8: 182-189.
- [17] Amresh, G., Paras, N. S. and Chandana, V.
 A. (2008). Toxicological screening of traditional medicine of laghupatha (*Cissampelos parara*) in experimental

animals. *Journal of Ethnopharmacology*, 116: 454-460.

- [18] Lorke, D. (1983). A new approach to practical acute toxicity. *Archives Toxicity*, 54: 278-287.
- [19] Oyedemi, S. O., Adewusi, E. A., Aiyegoro, O. A. and Akinpelu, D. A. (2011). Antidiabetic and Haematological Effect of Aqueous Extract of Stem Bark of *Afzelia africana* (Smith) on Streptozotocin-induced Diabetic Wistar rats. *Asian Pacific Journal of Tropical Biomedicine*, 20: 353-358.
- [20] Trease, G. E. and Evans, W. O. (2009). Trease and Evans Pharmacognosy, Sixteenth Edition. New York: Sauders Elsevier Limited, 104-262pp.
- [21] Adeneye, A. A., Ajagbona, O. P, Adeleke, T.
 I. and Bello, S. O. (2006). Preliminary toxicity and phytochemical studies of the stem bark aqueous extract of *Musanga cecropiodes* in rats. *Journal of Ethnopharmacology*, 105: 374-379.
- [22] Sofowora, A. (1996). Research on medicinal plants and traditional medicine in Africa. *Journal of Alternative and Complementary Medicine*. 2: 365-372.
- [23] Ogwumike, O. O. (2002). Haemopoietic effect of aqueous extract of leaf sheath of *Sorghum bicolor* in albino rats. *African Journal of Biomedical Research*, 5: 69-71.
- [24] Kubmarawa, D., Andenyang, I. F. H. and Magomya, A. M. (2009). Proximate composition and amino acid profile of two non-conventional leafy vegetables (*Hibiscus cannabinus* and *Haematostaphis barteri*). *African Journal of Food Science*, 3: 233-236.
- [25] Amin-Mir, M., Sawhney, S. S. and Jassal, M.
 M. S. (2013). qualitative and quantitative analysis of phytochemicals of *Taraxacum officinale*. Wudpecker Journal of Pharmacy and Pharmacology, 2 (1): 001-005.

- [26] Lewis, S. M., Bain, B. J. and Bates, I. (2001).
 Dacie and Lewis: Practical Haematology.
 Ninth Edition, London, New York: Elsevier, 9-40pp.
- [27] Esenowo, G. J., Sam, S. M., Bala, D. N., Ekpo,
 B. A. J. and Edung, E. M. (2010).
 Phytochemical screening and the haematological effect of *Peristrophe bicalculata* (Retz) nees diet preparation in albino rats. *World Journal of Applied Science and Technology*, 2(2): 277-281.
- [28] Farine, J. P., Legal, L., Moreteau, B. and Quee, J. (1996). Volatile components of ripe fruits of *Moinde citrifolia* and their effects on *Drosiphila*. *Journal of Phytochemistry*, 41: 433-438.
- [29] Latha, P. G. and Panikkar, K. R. (2000).
 Inhibition of Chemical Carcinogenesis in Mice by Mice by *Ixora coccinea* Flowers.
 Pharmaceutical Biology, 38 (2): 152-156.
- [30] Baba-Mousa, F., Akpagana, K. and Bouchet,
 P. (1999). Anti-fungal activities of seven
 West African Combretaceace used in
 Traditional Medicine. Journal of Ethnopharmacology, 66: 335-338.
- [31] Singh, A. P. (2006). Short Review: Distribution of steroid like compounds in Plant Flora. *Pharmacological Management*, 2(6): 87-89.
- [32] Szudelski, T. (2001) The mechanism of alloxan and streptozotocin action in β-cells of the rat pancreas. *Physiology Research*, *50*: 536-546.
- [33] Katzung, B. G. and Trevor, A. (2009). Masters SB basic and clinical pharmacology. Mc-Graw Hill; 346p.
- [34] Odoemena, C. S. I. Ekpo, B. A. J. and Luke, M. I. (2007). Hypoglycemic activity of Ocimum gratissimum (Linn) ethnaolic leaf extract in normoglycaemic and hyperglycaemic rats", J. Tropical Biosciences, 7: 24-28.

 [35] Schimizu, M., Ito, T., Rshima, S., Mayashi, T., Arisawa, M., Morita-Kurokowa, S. and Ito-Hasimato, Y. (1984). Inhibition of lens aldose reductase by flavonoids. *Phytochemistry*, 23: 1885-1888.