

### In vitro Studies on the Inhibition of $\alpha$ -amylase and $\alpha$ -glucosidase by Methanolic Extract of Barleria Prionitis L. and Psidium guajava

Akshay R. Yadav\*, Dr. Shrinivas K. Mohite

Department of Pharmaceutical Chemistry, Rajarambapu College of Pharmacy, Kasegaon, Dist- Sangli, Maharashtra, India-415404 \*Corresponding author E-mail: akshayyadav24197@gmail.com

#### ABSTRACT

The management of the blood glucose level is a critical strategy in the control of diabetes complications. There are many and diverse therapeutic strategies in the management of Type II diabetes. The inhibition of carbohydrate hydrolyzing enzymes such as  $\alpha$ -amylase can be an important strategy to lower postprandial blood glucose levels. Such inhibitors which find application in the clinical practice for management of diabetes are known to be associated with various gastrointestinal side effects. Therefore, it is the need of time to identify and explore the amylase inhibitors from natural sources having fewer side effects. In the present study, 50% volume per volume methanolic extracts of *Barleria Prionitis* and psidium guajava subjected to *in vitro* analysis of antidiabetic effect by alpha-amylase and alpha-glucosidase inhibitory activity of the methanolic extract of the plant individually against alpha-amylase enzyme and alpha-glucosidase enzyme were examined in different concentrations (3.90–500 µg/mL), where acarbose used as a positive control. The percentage inhibition of *Barleria Prionitis* showed the highest alpha-amylase and alpha-glucosidase inhibitory activity. Half-maximal inhibitory concentration value *Barleria Prionitis* was found for alpha-amylase and alpha-glucosidase inhibition. This study suggests that the methanolic extract of all two plants have antidiabetic property, among these three plants *Barleria Prionitis* showed potent enzyme inhibition as compared to other plant extracts and standard acarbose.

Keywords: Barleria Prionitis, Psidium guajava, Alpha-amylase, Alpha-glucosidase, Antidiabetic.

#### I. INTRODUCTION

Diabetes mellitus is a chronic multifactorial disorder and one of the non-communicable life-threatening metabolic diseases involving huge health-care cost and high mortality rate<sup>1</sup>. In 2015, it was found that it affecting 422 million adults globally. The majority of them were between 40 and 59 years and around 80% lived in middle- and low-income countries<sup>2</sup>. It was found that more than 4.9 million deaths were caused alone with diabetes and the number of diabetes patients will increase up to 55% by 2035, reaching 592 million aging between 20 and 79 years. These are non-infectious and non-transmissible. It is characterized by chronic hyperglycemia with disturbance of carbohydrate, fat, and protein metabolism due to the insufficient secretion of insulin by the pancreas and by the resistance to the action on insulin in various issues, i.e., muscle, liver, and adipose, which results in impaired uptake of glucose<sup>3-4</sup>. Postprandial hyperglycemia is one of the earliest observable abnormalities of glucose homeostasis, in which blood glucose level remains high after consuming meal and plays an important role in the development of type 2 diabetes and associated chronic complications, such as micro- and macro-vascular disorder. Management of plasma glucose levels is essential for delaying or preventing type-2 diabetes<sup>5</sup>. Insulin secretion through medication and/or dietary supervision, it is possible to reach this goal. Decreasing the postprandial glucose level is one of the therapeutic approaches for treating type-2 diabetes; for example slowing the glucose absorption through inhibition of the carbohydrates-hydrolyzing enzymes present in the small intestinal brush border,  $\alpha$ glucosidase, and  $\alpha$ -amylase. These are responsible for the breakdown of oligosaccharides and disaccharides into monosaccharides<sup>6-7</sup>. Fruits and vegetables that are consumed worldwide have excellent sources of bioactive compounds and having capacity reducing the risk of developing diabetes. Postprandial glucose levels can be regulated through  $\alpha$ -glucosidase inhibition. Inhibition of these enzymes delay and in some cases halt carbohydrate digestion, thus prolonging overall carbohydrate digestion time, causing a reduction in the rate of glucose absorption and consequently reducing postprandial plasma glucose rise Nowadays,  $\alpha$ -glycosidase inhibitors such as acarbose, miglitol, and voglibose are oral blood glucose-lowering drugs commonly used. They postprandial hyperglycemia decrease without inducing insulin secretion; these compounds do not induce hypoglycemia and have good safety; although, the gastrointestinal adverse effect may limit longterm compliance to therapy. Several medicinal plants species have been used to control diabetes in the traditional medicinal systems of many cultures worldwide. The potential role of medicinal plants as inhibitors of  $\alpha$ -amylase and  $\alpha$ -glucosidase has been reviewed by several authors. A variety of plants has been reported to show an enzymatic inhibitory activity, and so many are relevant to the treatment of type-2 diabetes. The research for a new group of agents from natural resources, especially from traditional medicine becomes an attractive approach for the treatment of postprandial hyperglycemia. It is revealed that there is a direct relationship between phenolic compounds, flavonoids, and tannins and the ability to inhibit  $\alpha$ -amylase and  $\alpha$ -glycosidase activities. These phenolic compounds have a positive effect on diabetes, by inhibiting the two keys enzymes hydrolyzing carbohydrates in the digestive tract<sup>8-9</sup>. The Soxhlet extraction, which is a standard technique, is a continuous solvent extraction method. Extraction systems are used to conduct routine solvent extractions of soils, sediments, sludge, polymers and plastics, pulp and paper, biological tissues, textiles and food samples<sup>10-25</sup>. Experiments have proved that microwaves, in comparison with the soxhlet extraction, use a lesser volume of solvent and sample and perform extraction at a much faster rate<sup>26-</sup> <sup>53</sup>. In the discovery of effective medicines for prevention and treatment, an outbreak of coronavirus disease (COVID-19) caused by the novel extreme acute respiratory syndrome coronavirus-2 (SARS-CoV-2) poses an unprecedented obstacle. The proximity to the patient during dental care, high generation of aerosols, and the identification of SARS-CoV-2 in saliva have suggested the oral cavity as a potential reservoir for COVID-19 transmission. Soon, someday, you might be making your own drugs at home. That is because researchers have adapted a 3D printer from basic, readily available medicinal active agents fed into a drug delivery system<sup>54-74</sup>.

#### **II. MATERIALS AND METHODS**

#### Chemicals and Reagents

Porcine pancreatic  $\alpha$ -amylase (PPA), 3,5dinitrosalicylic acid (DNS color reagent), Potassium phosphate buffer solution (PBS), p-Nitrophenyl- $\alpha$ -Dglucopyranoside (pNPG),  $\alpha$ -glucosidase, and ascorbose were purchased from Sigma Aldrich. Soluble starch potato, sodium potassium tartrate, sodium chloride, disodium hydrogen phosphate, and sodium hydroxide were from Merck Chemical Supplies (India). All the chemicals, including the solvents used in this study, were of analytical grade.

#### Plant Materials

The fresh matured leaves of the *B. prionitis and Psidium guajava* were collected randomly, from Sangli region, Maharashtra, India. Department of Botony, Yashwantrao Chavan College of Science, Karad has identified the plant and authenticated it.

#### Preparation of Plant Extract

Shade drying was done for almost a month to prevent sunlight chemical degradation. The dried material was grinded and transformed in coarse powder with the aid of a grinder. The extraction of *B. prionitis and Psidium guajava* with solvent methanol was carried out by microwave extraction, and excess solvent present was evaporated.

# In vitro methods employed in antidiabetic studies $\alpha$ -amylase inhibition activity

PPA (enzyme commission 3.2.1.1) solution was dissolved in 20 mM phosphate buffer (pH 6.9 with 6.7 mM sodium chloride) to give a concentration of 1 U/ml. Starch solution (1%, w/v) was obtained by stirring 0.1 g of potato starch in 100 ml of 20 mM of phosphate buffer (pH 6.9 with 6.7 mM sodium chloride) as a substrate. A total of 100  $\mu$ l of plant extract solution and 100 µL of the enzyme were preincubated at 37°C for 30 min. After preincubation 100  $\mu l$  of a 1% starch solution was added. The reaction mixtures were then incubated at 37°C for 20 min. The reaction was stopped with 200  $\mu$ L of DNS color reagent and placed in boiling water for 5 min and cooled to room temperature. Add 200 µl of reaction mixture into the 96-well microplate after diluted with 1.5 ml of distilled water. The  $\alpha$ -amylase activity was determined by measuring the absorbance of the mixture at 540 nm. Acarbose was used as positive control. Percentage inhibition was calculated by comparing against control optical density with the test group<sup>75-77</sup>.

#### $\alpha$ -glucosidase inhibitory activity

The  $\alpha$ -glucosidase inhibitory activity was performed with a set of microwell. The enzyme solution

containing 20  $\mu$ l  $\alpha$ -glucosidase (0.1 unit/ml) enzyme solutions were added in 96 microwell plate except blank well. A volume of 120  $\mu$ l 0.1 M PBS solutions were added into the well-containing enzyme and 140  $\mu$ l 0.1 M PBS in blank well and 160  $\mu$ l PBS in extract blank well. Ten microliters of test samples (Acarbose or test samples) were added into the enzyme solution in microplate wells and then incubated for 15 min at 37°C. Twenty microliters of pNPG solutions were added to the microwell plate and incubated the plate for 15 min at 37°C. The reaction was terminated by adding 80  $\mu$ l of 0.2 M sodium carbonate solution.

- Test solution contains: 20 µl enzyme + 120 µl PBS
  + 10 µl of test samples + 20 µl pNPG + 80 µl stop reagent.
- Control solution: All reaction mixture without test samples (20 µl enzyme + 130 µl PBS + 20 µl pNPG + 80 µl stop reagent).
- Blank solution: All reaction mixture except  $\alpha$ -glucosidase enzyme (140  $\mu$ l PBS + 10  $\mu$ l of test samples + 20  $\mu$ l pNPG + 80  $\mu$ l stop reagent)
- Extract blank solution: 10 μl extract + 160 μl PBS + 80 μl stop reagent.

The absorbance of the wells was measured with a microplate reader. at 405 nm, while the reaction system without plant extracts was used as control. The system without  $\alpha$ -glucosidase was used as blank, and acarbose was used as positive control. Each experiment was conducted in triplicate. The percentage enzyme inhibition and half-maximal inhibitory concentration (IC<sub>50</sub>) was calculated.

#### Calculation of half-maximal inhibitory concentration

The concentration of plant extracts required to scavenge 50% of the radicals (IC50) was calculated by using the percentage scavenging activities at five different concentrations of the extracts. Percentage inhibition (I%) was calculated by<sup>78-79</sup>

 $I\% = (A_c - A_s)/Ac \times 100$ 

Where,

 $A_c$  is the absorbance of the control and  $A_s$  is the absorbance of the sample.

#### III. RESULTS AND DISCUSSION

Antidiabetic plants have a major role in inhibiting the glucose level thus providing protection to human against hyperglycemia. Realizing the facts his research was carried out to evaluate the antidiabetic activity of methanolic extract of the selected plants. The *in vitro* antidiabetic activity of these plants extract was detected by measurement of glucose uptake in L6 cell lines.

#### $\alpha$ -Amylase inhibition activity

In this study, the *in vitro*  $\alpha$ -amylase inhibitory activities of the hydro-ethanolic extract of the *B. prionitis and Psidium guajava* was investigated. The results of the experiment showed that there was a dose-dependent increase in percentage inhibitory activity against  $\alpha$ -amylase enzyme [Table 1]. The IC<sub>50</sub> values were determined using potato starch (1%, w/v) in 20 mM phosphate buffer (pH 6.9 containing 6.7 mM sodium chloride) is used as substrate *(in vitro)* and tested sample concentration ranged from 3.90 to 500 µg/ml. *B. prionitis* extract showed highest  $\alpha$ -amylase inhibitory activity as compared to the standard drug (acarbose).

Table	1:	$\alpha$ -Amylase	inhibition	data	at	different
concer	itra	tion of test sa	mples			

Concentration	Percentage of inhibition			
(µg/ml)	Barleria	Psidium	Standard	
	Prionitis	guajava	(acarbose)	
3.90	42.28	34.23	14.34	
7.81	51.36	42.78	29.51	
15.63	60.12	55.21	34.82	
31.25	67.39	58.27	45.91	
62.50	75.10	60.16	58.18	
125.00	82.91	71.90	69.42	
250.00	94.82	84.91	80.49	
500.00	100.00	89.02	97.23	

dependent increase in percentage inhibitory activity against  $\alpha$ -glucosidase enzyme [Table 2]. methanolic extracts of the B. prionitis and Psidium guajava showed  $\alpha$ -glucosidase inhibitory potential. The halfmaximal inhibitory concentration values were determined using paranitrophenyl- $\alpha$ -D-glucopyranoside as substrate (in vitro) and tested sample concentration ranged from 9.30 to 500 µg/ml. B. prionitis extract showed highest  $\alpha$ -glucosidase inhibitory activity as compared to standard drug (acarbose). Inhibition of  $\alpha$ -amylase and  $\alpha$ -glucosidase enzymes involved in the digestion of carbohydrates, which can significantly decrease the postprandial increase of blood glucose after a mixed carbohydrate diet and therefore can be play an important role in the management of postprandial blood glucose level in type 2 diabetic patients and borderline patients. According to numerous in vitro studies, inhibition of  $\alpha$ -amylase and  $\alpha$ -glucosidase is believed to be one of the most effective approaches for diabetes care.

results of experiment showed that there was a dose-

**Table 2:**  $\alpha$ -Glucosidase inhibition data at different concentration of test samples

Concentration	Percentage of inhibition				
(µg/ml)	Barleria	Psidium	Standard		
	Prionitis	guajava	(acarbose)		
3.90	35.67	8.45	5.76		
7.81	40.38	22.79	29.54		
15.63	48.21	34.12	35.01		
31.25	54.91	44.61	44.12		
62.50	60.18	51.80	52.01		
125.00	74.89	60.35	58.39		
250.00	88.34	71.87	65.12		
500.00	94.12	78.32	71.43		

#### **IV. CONCLUSION**

## $\alpha$ -glucosidase inhibition activity

In this study, the *in vitro*  $\alpha$ -glucosidase inhibitory activities of the hydro-alcoholic extract of the *B. prionitis and Psidium guajava* was investigated. The

Conventionally, many herbal formulations are using as single herb or in combinations of several different herbs. It believed that poly herbs show synergistic effect. The herbal formulation includes either plant raw material or plant extracts. Here, all selected plants are collected from the Chambal Valley of India to investigate the antidiabetic properties. This study provides the evidence that 50% v/v methanolic extracts of all two plants *B. prionitis and Psidium guajava* are having potent enzyme inhibitory actions which are responsible for hyperglycemia. However, more efforts are needed for the isolation and characterization of bioactive compounds and further evaluation of biological properties.

#### V. REFERENCES

- [1]. Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. J Ethnopharmacol 2002; 81: 81-100.
- [2]. Bnouham M, Ziyyat A, Mekhfi H, Tahri A, Legsseyer A. Medicinal plants with potential antidiabetic activity – A review of ten years of herbal medicine research (1990-2000). Int J Diabetes Metab 2006; 14: 1-25.
- [3]. Mentreddy SR. Medicinal plant species with potential antidiabetic properties. J Sci Food Agric 2007; 87: 743-50.
- [4]. Benalla W, Bellahcen S, Bnouham M. Antidiabetic medicinal plants as a source of alpha glucosidase inhibitors. Curr Diabetes Rev 2010; 6: 247-54.
- [5]. Ponnusamy S, Zinjarde SS, Bhargava SY, Kumar AR. Potent α-amylase inhibitory activity of Indian ayurvedic medicinal plants. BMC Complement Altern Med. 2011; 11: 5.
- [6]. Tadera K, Minami Y, Takamatsu K, Matsuoka T.
  Inhibition of alpha-glucosidase and alpha-amylase by flavonoids. J Nutr Sci Vitaminol (Tokyo). 2006; 52: 149-53.
- [7]. Adisakwattana S, Chanathong B.
   Alpha-glucosidase inhibitory activity and lipid-lowering mechanisms of Moringa oleifera leaf extract. Eur Rev Med Pharmacol Sci. 2011; 15: 803-8.

- [8]. Lo Piparo E, Scheib H, Frei N, Williamson G, Grigorov M, Chou CJ. Flavonoids for controlling starch digestion: Structural requirements for inhibiting human alpha-amylase. J Med Chem 2008;51:3555-61.
- [9]. Rubilar M, Jara C, Poo Y, Acevedo F, Gutierrez C, Sineiro J, et al. Extracts of maqui (Aristotelia chilensis) and murta (Ugni molinae turcz): Sources of antioxidant compounds and α-glucosidase/α-amylase inhibitors. J Agric Food Chem. 2011; 59: 1630-7.
- [10].Yadav A, Mohite S, Magdum C. Synthesis, Characterization and Biological Evaluation of Some Novel 1,3,4-Oxadiazole Derivatives as Potential Anticancer Agents. Int. j. sci. res. sci. technol. 2020; 7(2): 275-282.
- [11].Yadav A, Mohite S. Anticancer Activity and In-Silico ADMET Analysis of Malvastrum Coromandelianum. International Journal of Pharma Sciences and Research. 2020; 11(5): 71-73.
- [12].Yadav A, Mohite S. Cancer- A Silent Killer: An Overview. Asian J. Pharm. Res. 2020; 10(3): 213-216.
- [13].Chitruk A, Yadav A, Rode P, Mohite S, Magdum C. Synthesis and toxicological evaluation using brine shrimp lethality assay of Novel 1,2,4triazole derivatives with anticancer activity. Int. J. Curr. Adv. Res. 2020; 09(08)(A): 22877-22881.
- [14].Yadav A, Mohite S. Design, Synthesis and Characterization of Some Novel benzamide derivatives and it's Pharmacological Screening. Int. j. sci. res. sci. technol. 2020; 7(2): 68-74.
- [15].Honmane P, Yadav A, Singh S, Mohite S. Microwave Assisted Synthesis of Novel Benzimidazole Derivatives as Potent Antileishmanial and Antimalarial Agents. Int. J. Curr. Adv. Res. 2020; 09(07)(B): 22742-22746.
- [16].Yadav A, Mohite S. A Brief Review: Microwave Chemistry and its Applications. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(3): 191-197.

- [17].Chitruk A, Yadav A, Rode P, Mohite S, Magdum C. Microwave assisted synthesis, antimicrobial and anti-inflammatory potential of some novel 1,2,4-triazole derivatives. Int. j. sci. res. sci. technol. 2020; 7(4): 360-367.
- [18].Yadav A, Mohite S. Antioxidant Activity of Malvastrum Coromandelianum Leaf extracts. Research J. Topical and Cosmetic Sci. 2020; 11(2): 59-61.
- [19].Yadav A, Mohite S. In-Silico ADME Analysis of 1, 3, 4-oxadiazole derivatives as CDK9 Inhibitors. International Journal of Chemical Science. 2020; 4(3): 01-04
- [20].Yadav A, Patil S, Dharanguttikar V, Mohite S. Anthelmintic Activity of Malvastrum Coromandelianum Leaf Extracts against Pheretima Posthuma and Ascardia Galli. Int J Sci Res Chemi. 2020; 5(6): 18-24.
- [21].Yadav A, Mohite S. Formulation and Evaluation of Antidandruff Shampoo. Research J. Topical and Cosmetic Sci. 2020; 11(2): 55-58.
- [22].Yadav A, Mohite S, Magdum C. Comparative Study of Conventional and Microwave Assisted Synthesis of some Organic Reactions. Asian J. Pharm. Res. 2020; 10(3): 217-220.
- [23].Yadav A, Mohite S. Different Techniques and Characterization of Polymorphism with their Evaluation: A Review. Asian J. Pharm. Tech. 2020; 10(3): 213-216.
- [24].Yadav A, Mohite S. Aquasomes as a Self Assembling Nanobiopharmaceutical Carrier System for Bio-Active Molecules. Research J. Topical and Cosmetic Sci. 2020; 11(2): 66-70.
- [25].Yadav A, Mohite S. Anthelmintic and Antibacterial Activity of Psidium Guajava Leaf Extracts. Int J Sci Res Chemi. 2020; 5(6): 06-11.
- [26].Yadav A, Dange V, Mohite S. Pathogensis of Cell Injury. Int J Sci Res Chemi. 2020; 5(6): 12-18.
- [27].Suryawanshi V, Yadav A, Birajdar R, Jagtap N, Vambhurkar G, Patil P. Optimization of ayurvedic herbal medicine by nanoformulation. Asian J. Res. Pharm. Sci. 2019; 9(1): 55-56.

- [28].Yadav A, Honmane P, Bhosale M, Chitruk A, Rode P, Birajdar R, Rajput M, Suryawanshi V, Patil S, Patil, Jagtap N, Mohite S, Dange V, Vambhurkar G. Antifungal Activity of Malvastrum Coromandelianum Leaf Extracts. International Journal of Scientific Research in Chemistry. 2020; 5(6): 01-05.
- [29].Yadav A, Mohite S. Potential Role of Peptides for Development of Cosmeceutical skin Product. Research J. Topical and Cosmetic Sci. 2020; 11(2): 77-82.
- [30].Yadav A, Mohite S. Green Chemistry approach for Microwave assisted synthesis of some Traditional Reactions. Asian J. Research Chem. 2020; 13(4): 261-264.
- [31].Yadav A, Mohite S. Screening of In-vitro antiinflammatory and Antifungal assay of Psidium guajava Leaf Extracts. Research J. Topical and Cosmetic Sci. 2020; 11(2): 62-64.
- [32].Yadav A, Mohite S, Magdum C. Microwave assisted synthesis of some Traditional reactions: Green chemistry approach. Asian J. Research Chem. 2020; 13(4): 275-278.
- [33].Yadav A, Mohite S. Applications of Nanotechnology in Cosmeceuticals. Research J. Topical and Cosmetic Sci. 2020; 11(2): 83-88.
- [34].Yadav A, Mohite S. Rajput M, Suryawanshi V, Birajdar R, Patil M. Antioxidant Activity of Psidium guajava Leaf Extracts. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(3): 159-161.
- [35].Yadav A, Mohite S. ADME analysis of phytochemical constituents of Psidium guajava. Asian J. Res. Chem. 2020; 13(5): 373-375.
- [36].Rode P, Yadav A, Chitruk A, Mohite S, Magdum C. Synthesis, Anticancer and Molecular Docking Studies of N-(1H-benzimidazol-2-yl-carbamothioyl)benzamide Analogues. International Journal of Scientific Research in Science and Technology. 2020; 5(6): 204-212.
- [37].Yadav A, Mohite S. Recent advances in protein and peptide drug delivery. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(3): 205-212.

International Journal of Scientific Research in Chemistry (www.ijsrch.com) | Volume 5 | Issue 1

- [38].Yadav A, Mohite S. A Novel approach for treatment of COVID-19 with Convalescent Plasma. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(3): 227-230.
- [39].Yadav A, Mohite S. A Review on Novel Coronavirus (COVID-19). International Journal of Pharma Sciences and Research. 2020; 11(5): 74-76.
- [40].Yadav A, Mohite S. A Review on severe acute respiratory infection (SARI) and its clinical management in suspect/confirmed novel coronavirus (nCoV) cases Res. J. Pharma. Dosage Forms and Tech. 2020; 12(3): 178-180.
- [41].Yadav A, Mohite S. A Review on Zika Virus Infection. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(4): 245-249.
- [42].Honmane P, Yadav A, Singh S, Mohite S. Formulation and Evaluation of Herbal Ointment Containing Eclipta Alba (L.) Extract. Seybold Rep. 2020; 25(10): 569-577.
- [43].Yadav A, Mohite S. Toxicological Evaluation of Psidium guajava Leaf Extracts using Brine Shrimp (Artemia salina L.) Model. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(4): 198-120.
- [44].Honmane P, Yadav A, Singh S, Mohite S. Synthesis, Characterization and Antiplatelet Activity of Antithrombotic novel 2,5-substituted aryl-7-phenyl-1,3,4-oxadiazolo-3,2-a]-1,3,5triazine Derivatives. Journal of University of Shanghai for Science and Technology. 2020; 22(11): 881-898.
- [45].Patil S, Yadav A, Chopade A, Mohite S. Design, Development and Evaluation of Herbal Mouthwash for Antibacterial Potency against Oral Bacteria. Journal of University of Shanghai for Science and Technology. 2020; 22(11): 881-898.1137-1148.
- [46].Yadav A, Mohite S. Homology Modeling and Generation of 3D-structure of Protein. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(4): 218-224.

- [47].Honmane P, Yadav A, Singh S, Mohite S. Synthesis of Pyrazole Acrylic acid based Oxadiazole and Amide Derivatives as Larvicidal and Antitubercular agents. Seybold Rep. 2020; 25(10): 516-530.
- [48].Yadav A, Mohite S. Recent Advances in the Ultrasound-Assisted Synthesis of Oxadiazole and Thiazole Derivatives. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(4): 225-228.
- [49].Yadav A, Mohite S. An Overview on Ebola Virus Disease. Res. J. Pharma. Dosage Forms and Tech.2020; 12(4): 230-235.
- [50].Yadav A, Mohite S. Carbon Nanotubes as an Effective Solution for Cancer Therapy. Res. J. Pharma. Dosage Forms and Tech. 2020; 12(4): 238-241.
- [51].Honmane P, Yadav A, Singh S, Mohite S. 3D printing technology in pharmaceuticals and biomedical. World J Pharm Pharm Sci. 2020; 9(9): 598-609
- [52].Yadav A, Mohite S, Magdum C. Preparation and Evaluation of Antibacterial Herbal Mouthwash against Oral Pathogens. Asian J. Res. Pharm. Sci. 2020; 10(3): 149-152.
- [53].Gavali K, Yadav A, Howal R, Tamboli A. Preliminary Phytochemical Screening and HPTLC Finger printing of Leaf Extracts of Tectona grandis Linn Journal of University of Shanghai for Science and Technology. 2020; 22(11): 1804-1815.
- [54].Rajput M. D, Yadav A. R, Mohite S. K. Synthesis, Characterization of Benzimidazole Derivatives as Potent Antimicrobial Agents. International Journal of Pharmacy & Pharmaceutical Research. 2020; 17(4): 279-285.
- [55].Dange V, Dinde S, Doiphode A, Dhavane S, Dudhal B, Shid S, Yadav A. Formulation and Evaluation of Herbal gel Containing Lantana Camara for Management of Acne Vulgaris. Journal of University of Shanghai for Science and Technology.2020; 22(11): 799-809.

- [56].Yadav A, Mohite S. Screening of In-vitro antiinflammatory and Antibacterial assay of Malvastrum Coromandelianum. International Journal of Pharma Sciences and Research. 2020; 11(4): 68-70.
- [57].Rajput M, Yadav A. Green Chemistry Approach for Synthesis of Some 1,3,4-Oxadiazole Derivatives As Potent Antimalarial Agents Journal of University of Shanghai for Science and Technology. 2020; 22(11): 1854-1869.
- [58].Suryawanshi V, Yadav A, Mohite S. Toxicological Assessment using Brine Shrimp Lethality Assay and Antimicrobial activity of Capparis Grandis. Journal of University of Shanghai for Science and Technology. 2020; 22(11): 746-759.
- [59].Pathade K, Mohite S, Yadav A. 3D-QSAR And ADMET Prediction Of Triazine Derivatives For Designing Potent Anticancer Agents. Journal of University of Shanghai for Science and Technology. 2020; 22(11): 1816-1833.
- [60].Rode P, Yadav A, Chitruk A, Mohite S, Magdum C. Microwave assisted synthesis, toxicological assessment using brine shrimp lethality assay and antimicrobial potential of new series of benzimidazole derivatives. Int. J. Curr. Adv. Res. 2020; 09(08)(A): 22900-22905.
- [61].Bhosale M, Yadav A, Magdum C, Mohite S. Molecular Docking Studies, Synthesis, Toxicological Evaluation using Brine Shrimp (Artemia salina L.) Model and Anti-inflammatory Activity of Some N-(substituted)-5-phenyl-1,3,4thiadiazol-2-amine Derivatives. Int J Sci Res Sci & Technol. 2020; 7(5): 51-62.
- [62].Jagtap N, Yadav A, Mohite S. Synthesis, Molecular Docking Studies and Anticancer Activity of 1,3,4-Oxadiazole-3(2H)-thione Derivatives. Journal of University of Shanghai for Science and Technology.2020; 22(11):535-550.
- [63].Bhosale M, Yadav A, Magdum C, Mohite S. Microwave Assisted Synthesis, Molecular Docking Studies and Anticancer Screening of Some 1,3,4-thiadiazole Derivatives. Journal of

University of Shanghai for Science and Technology.2020; 22(11):520-534.

- [64].Birajdar R, Yadav A, Patil S, Chitruk A, Kane S, Mohite S, Magdum C. Pharmacognostic and Phytochemical Investigation, Molecular Docking Studies of Phytoconstituents and Anticancer Potential of Capparis Decidua (Forsk) Edgew. Journal of University of Shanghai for Science and Technology. 2020; 22(11): 500-519.
- [65].Yadav A, Mohite S. Pharmacophore Mapping and Virtual Screening. Int J Sci Res Chemi. 5(5): 77-80.
- [66].Yadav A, Mohite S. Photochemistry and Spectroscopy. Int J Sci Res Chemi. 5(5): 71-76.
- [67].Yadav A, Mohite S. Pharmaceutical Process Scale-Up. Int J Sci Res Chemi. 5(5): 49-55.
- [68].Yadav A, Mohite S. Transforming Global Health. Int J Sci Res Chemi. 5(6): 41-48.
- [69].Yadav A, Mohite S. FDA Lifecycle Approach to Process Validation. Int J Sci Res Chemi. 5(6): 35-40.
- [70].Yadav A, Mohite S. Mass Transfer Effects and Performance of Immobilized Enzymes. Int J Sci Res Chemi. 5(5): 51-56.
- [71].Yadav A, Mohite S. Phytochemical and Pharmacological Review of Embelia ribes. Int J Sci Res Chemi. 5(5): 57-62.
- [72].Pawara N, Yadav A, Mohite S. Pharmacognostic, Phytochemical Investigation and Antioxidant Potential of Embelia ribes. Int J Sci Res Chemi. 5(6): 27-34.
- [73].Yadav A, Rajput M, Gavali K, Mohite S. In-vitro Hypoglycemic Activity of Barleria prionitis L. Int J Sci Res Chemi. 5(5): 63-70.
- [74].Bhosale M, Yadav A, Magdum C, Mohite S. Synthesis, molecular docking studies and biological evaluation of 1,3,4-thiadiazole derivatives as antimicrobial agents. Int. J. Curr. Adv. Res. 2020; 09(08)(A): 22894-22899.
- [75].achan AK, Rao CV, Sachan NK. Extraction and evaluation of hypoglycemic and wound healing

potential of hydro-ethanolic extract of Alhagi pseudalhagi wild. Am J Ethnomed 2017; 4: 1-5.

- [76].Evans WC. Trease and Evans: Pharmacognosy. 15th ed. Edinburgh: Saunders/ Elsevier; 2005.
- [77].Doughari JH. Phytochemicals: Extraction methods, basic structures and mode of action as potential chemotherapeutic agents. In: Rao DV, editor. Phytochemicals – A Global Perspective of Their Role in Nutrition and Health. Croatia: InTech Open; 2012.
- [78].Harbone JB. Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. London: Chapman and Hall; 1998.
- [79].Kim KT, Rioux LE, Turgeon SL. Alpha-amylase and alpha-glucosidase inhibitionis differentially modulated by fucoidan obtained from Fucus vesiculosus and Ascophyllum nodosum. Phytochemistry 2013; 98: 27-33.

**Cite this article as :** Akshay R. Yadav, Dr. Shrinivas K. Mohite , "In vitro Studies on the Inhibition of ?-amylase and ?-glucosidase by Methanolic Extract of Barleria Prionitis L. and Psidium guajava ", International Journal of Scientific Research in Chemistry (IJSRCH), ISSN : 2456-8457, Volume 5 Issue 1, pp. 50-58, January-February 2020.

URL : http://ijsrch.com/IJSRCH120529